





Genome Sequence of *Corynebacterium pseudotuberculosis* Strain PA02 Isolated from an Ovine Host in the Amazon

Gabriel R. S. Muge,^a Adonney A. O. Veras,^a Pablo H. C. G. de Sá,^a Ana Lídia Queiroz Cavalcante,^a Jorianne Thyeska Castro Alves,^a Ezequiel Morais,^b André G. M. Silva,^b Luís C. Guimarães,^a Vasco Azevedo,^c Adriana Ribeiro Carneiro Folador,^a Artur Silva,^a Rommel T. J. Ramos^a

Center of Genomics and System Biology, Federal University of Pará (UFPA), Belém, Pará, Brazila; Institute of Biological Sciences, Federal University of Minas Gerais (UFMG), Belo Horizonte, Minas Gerais, Brazila; Institute of Veterinary Medicine, Federal University of Pará (UFPA), Campus of Castanhal, Pará, Brazila

In this work, we report the complete genome sequence of *Corynebacterium pseudotuberculosis* strain PA02 isolated from an ovine host. The genome contains 2,328,435 bp, a 52.2% G+C content, 2,035 coding sequences, 12 rRNA operons, 45 tRNAs, and 14 predicted pseudogenes.

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Address correspondence to Rommel T. J. Ramos, rommelthiago@gmail.com.

corynebacterium pseudotuberculosis is a Gram-positive, facultative intracellular, pleomorphic, nonsporulating, noncapsulated, nonmotile pathogen (1–3). It is the causative agent of caseous lymphadenitis (CLA) in small ruminants, which is found in all of the world's major sheep and goat herd areas and is characterized by the presence of caseous necrosis on the lymphatic glands or abscess formation in superficial lymph nodes and subcutaneous tissues. *C. pseudotuberculosis* also causes pyogranulomatous reactions, ulcerative lymphangitis, as well as mastitis and necrotic and ulcerative dermatitis in cattle (diseases with medical and veterinary relevance) (4).

CLA is a widespread disease that has been reported in many countries, including Australia, Brazil, Canada, New Zealand, South Africa, and the United States (4, 5). CLA leads to economic losses for sheep and goat farmers by causing skin deterioration and reducing yields of milk and wool. Moreover, *C. pseudotuberculosis* causes a visceral form of the disease that can affect internal organs, causing weight loss, death, and carcass condemnation (4).

C. pseudotuberculosis may be called the "perfect parasite" due to its ability to evade the immune system with apparent ease, once established within the host. *C. pseudotuberculosis* has a gene set repertoire for long-term survival outside a host environment (1).

C. pseudotuberculosis strain PA02 was isolated from an ovine host in Pará, Brazil. The genome sequencing was performed by Ion Torrent Personal Genome Machine (PGM) platform (Thermo, Fisher) using a fragment library, which produced a total of 392,463,062 bp representing a coverage of $171\times$. The FastQC version 0.11.4 software (http://www.bioinformatics.babraham.ac.uk/projects/fastqc) was used for assessing the quality of the raw data, and the FASTX-Toolkit (http://hannonlab.cshl.edu/fastx_toolkit) was used to trim and discard the reads with a Phred quality score below 20. The SPAdes genome assembler software (6) was used to generate 8 contigs, with an N_{50} of 713,155 bp, a maximum contig length of 819,241 bp, and a total size of 2,328,435 bp. The automatic genome annotation was performed

using RAST version 2.0 (7). tRNAs and rRNAs were predicted using tRNAScan-SE version 1.12 (8) and RNAmmer version 1.2 (9) software, respectively.

The manual curation of the annotation was performed through CLC Genomics Workbench version 8, Artemis version 16.0.0 (10), and the UniProt (http://www.uniprot.org) and NCBI nonredundant (https://www.ncbi.nlm.nih.gov) databases.

The *C. pseudotuberculosis* strain PA02 genome consists of a circular chromosome of 2,328,435 bp, with 52.2% G+C content, 2,035 coding sequences, 12 rRNAs, 45 tRNAs, and 14 pseudogenes.

Accession number(s). The *C. pseudotuberculosis* PA02 wholegenome shotgun project has the project accession number PRJNA318798. This version used for this project has the accession number CP015309.

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REFERENCES

- 1. Baird GJ, Fontaine MC. 2007. *Corynebacterium pseudotuberculosis* and its role in ovine caseous lymphadenitis. J Comp Pathol 137:179–210. http://dx.doi.org/10.1016/j.jcpa.2007.07.002.
- Dorella FA, Pacheco LG, Oliveira SC, Miyoshi A, Azevedo V. 2006. Corynebacterium pseudotuberculosis: microbiology, biochemical properties, pathogenesis and molecular studies of virulence. Vet Res 37:201–218. http://dx.doi.org/10.1051/vetres:2005056.
- Yeruham I, Elad D, Friedman S, Perl S. 2003. Corynebacterium pseudotuberculosis infection in Israeli dairy cattle. Epidemiol Infect 131:947–955. http://dx.doi.org/10.1017/S095026880300894X.
- 4. Soares SC, Silva A, Trost E, Blom J, Ramos R, Carneiro A, Ali A, Santos AR, Pinto AC, Diniz C, Barbosa EG, Dorella FA, Aburjaile F, Rocha FS, Nascimento KK, Guimarães LC, Almeida S, Hassan SS, Bakhtiar SM, Pereira UP, Abreu VAC, Schneider MPC, Miyoshi A, Tauch A, Azevedo

- V. 2013. The pan-genome of the animal pathogen *Corynebacterium pseudotuberculosis* reveals differences in genome plasticity between the biovar *ovis* and *equi* strains. PLoS One 8:e53818. http://dx.doi.org/10.1371/journal.pone.0053818.
- Araújo CL, Dias LM, Veras AA, Alves JT, Cavalcante AL, Dowson CG, Azevedo V, Ramos RT, Silva A, Carneiro AR. 2016. Whole-genome sequence of *Corynebacterium pseudotuberculosis* 262 biovar *equi* isolated from cow milk 4:4–5. http://dx.doi.org/10.1128/genomeA.00176-16.
- Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. J Comput Biol 19:455–477. http://dx.doi.org/10.1089/cmb.2012.0021.
- 7. Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formsma K, Gerdes S, Glass EM, Kubal M, Meyer F, Olsen GJ, Olson

- R, Osterman AL, Overbeek RA, McNeil LK, Paarmann D, Paczian T, Parrello B, Pusch GD, Reich C, Stevens R, Vassieva O, Vonstein V, Wilke A, Zagnitko O. 2008. The RAST server: Rapid Annotations using Subsystems Technology. BMC Genomics 9:75. http://dx.doi.org/10.1186/1471-2164-9-75.
- Lowe TM, Eddy SR. 1997. tRNAscan-SE: a program for improved detection of transfer RNA genes in genomic sequence. Nucleic Acids Res 25: 955–964. http://dx.doi.org/10.1093/nar/25.5.0955.
- Lagesen K, Hallin P, Rødland EA, Staerfeldt H-H, Rognes T, Ussery DW. 2007. RNAmmer: consistent and rapid annotation of ribosomal RNA genes. Nucleic Acids Res 35:3100–3108. http://dx.doi.org/10.1093/ nar/gkm160.
- 10. Rutherford K, Parkhill J, Crook J, Horsnell T, Rice P, Rajandream MA, Barrell B. 2000. Artemis: sequence visualization and annotation. Bioinformatics 16:944–945. http://dx.doi.org/10.1093/bioinformatics/16.10.944.